

THE QUALITY OF SEAHORSE JUVENILES *Hippocampus barbouri* AFTER MODIFYING NATURAL FEED *Artemia* NAUPLII TO *Phronima* sp.

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ABSTRACT

This study aims to determine the substitution rate of *Artemia* naupli to *Phronima* sp., which produces high-quality seahorses *H. Barbouri* juveniles. This research was carried out using CRD (Completed Randomized Design) with five treatments and three replications. The treatments were natural feeding with different levels of substitution, (A) 100% *Artemia* nauplii, (B) 75% *Artemia* nauplii+25% *Phronima* sp. (C) 50% *Artemia* nauplii +50% *Phronima*, (D) 25% *Artemia* nauplii+75% *Phronima* sp. and (E) 100% *Phronima* sp. Data were analyzed with Analysis of Variance and W-Tuckey test to determine the significant difference between the treatments. The parameters of the study included the analysis of feed quality (proximate and amino acid analysis) and survival. Results of the ANOVA indicated that the substitution of *Artemia* nauplii to *Phronima* sp. had a significant effect ($P < 0.05$) on survival rate. Based on the result of the proximate analysis of *Phronima*, the protein and fat content is insufficient for juvenile requirements, which only ranged from 37,12% and 3,82%. The results of the amino acid analysis of *Phronima*, the total of essential amino acids are deficient in juvenile requirements. The treatments of C and D produces the highest survival value of 96,67%. While in E treatment produce the lowest survival value of 63,33%. Based on the juvenile quality analysis, it can be concluded that the use of *Phronima* as an alternative feed for seahorse juveniles *H. barbouri* can be given at a substitution rate of up to 75%.

Keywords: *Artemia* naupli, *Hippocampus barbouri*, Juvenile, *Phronima* sp., Sea horse

INTRODUCTION

Seahorses are fish species that have high economic value and have been widely traded as ornamental fish, souvenirs, and dried as a raw material for traditional Chinese medicines (Rosa *et al.*, 2011). At present, the seahorse population has decreased due not only to habitat loss but also due to over-exploitation (Koldewey & Smith, 2010). Cultivation is the right answer to avoid overfishing and optimal use of resources. Some countries are known to have carried out seaweed hatchery activities, among others; the Philippines, Vietnam, China, India, and Australia. Whereas in Indonesia, the development of seaweed hatchery activities in both the mass and the backyard scale has been carried out in Lampung Sea Cultivation Center and also in South Sulawesi on the island of Sabutung of Pangkep Regency and Badi Island, Mattiro Deceng Village of Pangkep Regency.

Sea horses are passive predators, animals that wait for feed to pass and attack their prey by sucking using a long snout. So far, the type of natural feed given by seahorse juvenile is *Artemia salina* naupli, because, in addition to being natural in the culture process, it is also available commercially (Payne & Rippingale, 2000). However, *Artemia salina* is one of the natural feed with high selling

prices in the market. The quality requirements for feed use are continuity of availability, nutritional content, affordable prices, and not as a carrier of the disease (Handajani & Widodo, 2010). Therefore there's the need for alternative feed that can replace *Artemia salina*.

Santoso (2014) found the main feed derived from the Amphipoda order, complementary originating from the Mysida and Calanoid orders and supplementary derived from Harpatocoida, Isopoda and Euphasiacea order. *Phronima* sp. is a type of micro crustacea from the Amphipoda order which found naturally live in the waters of Waringtasi and Tasiwalie Villages of Pinrang Regency, South Sulawesi. *Phronima* has enormous potential as an alternative feed to replace *Artemia*, which is commonly used as feed on fish and shrimp seeds (Fattah *et al.*, 2014). The existence of *Phronima* sp. in the ponds of tiger shrimp pond maintenance, has been known to have succeeded in increasing the survival value by 70% compared with no *Phronima* maintenance,. The survival was only 10% (Fattah & Senong, 2008).

Based on the description above, information on *Phronima* sp. being as feed has never been done before for the *H. barbouri*'s seahorse juvenile rearing which is expected to be able to substitute

Artemia naupli which is still imported from the outside, so the need for research on the use of *Phronima* sp. as a sea horse juveniles feed. This study aimed to determine the level of substitution of *Phronima* in replacing *Artemia* naupli as an alternative feed for the rearing of *H. barbouri* juveniles so that it will produce the best survival.

MATERIALS AND METHODS

Test animals

The test animals used were 7-day-old *H. barbouri* seahorse juveniles with a body length of 1.5 cm. Juveniles were obtained from the spawning natural in the backyard. Seahorses broodstock obtained from Tanakeke waters, Takalar Regency, South Sulawesi.

Experimental container

Juvenile sea horses were maintained using 5-liter plastic containers equipped with buoys on each side. The container for juvenile sea horse designed by using a recirculation and water bath system equipped with filters (Figure 1). The purpose of this design was that the quality of water in the juvenile media remains stable and easy to control.

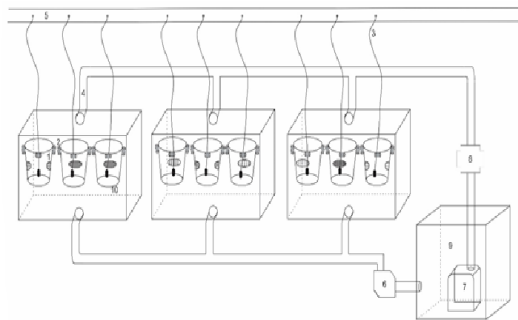


Figure 1. Design of a media container for *H. barbouri* juveniles

Notes: (1) juveniles media containers, (2) buoys, (3) aeration hoses, (4) inlet pipes, (5) oxygen distribution pipes, (6) biological filters, (7) pumps, (8) filter cartridges, (9) filtration containers and (10) filter.

Feed preparation

Artemia obtained from hatchery while *Phronima* sp. obtained from Tasiwalie Village, Pinrang then cultured in a 1-ton tub. Before the experiments started, the juveniles fasted a day, on the sixth day of rearing, which aimed to empty the contents of the previous feed of the juveniles. The treatments started on the seventh day (± 0.65 mm). This was due to the size of *Phronima* (± 0.45 mm), which was higher than the size of the juvenile mouth opening (± 0.4 mm). While the size of *Artemia* naupli was ± 0.3 mm. On the first until the fifth

day of rearing the juveniles were fed with the newly hatched *Artemia* naupli.

Broodstock of Sea horses

The broodstock spawned in the backyard was obtained from the catches of fishermen in the Tanakeke Islands, Takalar Regency, with lengths for male and ± 14 cm female. The broodstock was reared in a 700l tank. During rearing; the broodstock was given *Phronima* sp., the frequency of feeding twice as satiation, in the morning at 08.00 and in the afternoon at 16.00.

Juveniles rearing

Juveniles obtained from spawning then moved into the treatment container. The rearing of juveniles was 21 days with a density of 2 fish / L (Mulyadi, 2004) using containers with a volume of 5 liters of water. During research, water change was carried out as much as 50% every week and diluting the remaining feces every day on the bottom of the media container.

Experimental design and data analysis

This study was conducted using a completely randomized design with five treatments and three replications. The treatments were feeding with each different level density. On the first day until the fifth day of rearing was given the newly hatched *Artemia* naupli. Feeding for each treatment began on the seventh day of the study. (A) 100% *Artemia* naupli (as a control), (B) 75% *Artemia* naupli, 25% *Phronima* sp.; (C) 50% *Artemia* naupli, 50% *Phronima* sp. (D) 25% *Artemia* naupli, 75% *Phronima* sp. and (E) 100% *Phronima* sp.

The parameters of the study included testing the quality of feed by conducting proximate and amino acid analysis tests and calculation of survival rate. Data on feed quality testing will be described descriptively while the data from the calculation of survival obtained were analyzed using variance analysis. Data that had a significant effect ($P < 0.05$) were carried out by the W-Tuckey further test to see the differences in each treatment performed. Moreover, water quality measurements were also carried out during the research.

RESULTS AND DISCUSSION

Feed quality analysis

The proximate analysis results in (Table 1) produce the protein content values of treatments A, B, C, D and E which were 61.47%; 55.38%; 49.30%; 43.21% and 37.12%. The fat content produced in treatment A, B, C, D, and E were

21.55%; 17.12%; 12.69%; 8.25% and 3.82%. The value of crude fiber content in each treatment A, B, C, D, and E were 2.79%; 9.36%; 15.94%; 22.51% and 29.08%. The ash content in treatments A, B, C, D and E were 6.27%; 11.73%; 17.20%; 22.66% and 28.12%. NFE (Nitrogen Free Extract) or carbohydrates content in each treatment A, B, C, D and E which were 7.92%; 6.41%; 4.90%; 3.38% and 1.87%.

The total feed energy produced in all treatment A, B, C, D, and E was lower with increasing percentage substitution of *Artemia* naupli with *Phronima* sp. The total value of energy produced in treatments A, B, C, D, and E were 409.5 kcal/g respectively; 348.52 kcal/g; 287.58 kcal/g; 226.51 kcal/g and 165.53 kcal/g.

Table 1. The proximate analysis

The proximate composition	feed	Treatments				
		A	B	C	D	E
Crude protein (%)		61.47	55.38	49.30	43.21	37.12
Lipid (%)		21.55	17.12	12.69	8.25	3.82
Crude fiber (%)		2.79	9.36	15.94	22.51	29.08
Ash (%)		6.27	11.73	17.20	22.66	28.12
NFE (%)		7.92	6.41	4.90	3.38	1.87
DE (kkal)		409.5	348.52	287.58	226.51	165.53
CP Ratio (kkal/g protein)		6.66	6.29	5.83	5.24	4.46

Notes :

- 1.% dry weight;
2. A (100% *Artemia* naupli), B (75% *Artemia* + 25% *Phronima* naupli), C (50% *Artemia* + 50% *Phronima* naupli), D (25% *Artemia* + 75% *Phronima* naupli) and E (100% *Phronima*);
3. The value of the proximate analysis of treatments A and E is obtained from the proximate analysis results;
4. The proximate analysis of treatment B, C and D is obtained from the calculation;
5. NFE (Nitrogen Free Extract);
6. DE (Digestible Energy);
7. EP (Comparison of energy with protein);
8. Energy calculation based on NRC (1993) (1g protein: 3.5 kcal / g, 1g fat: 8.1 kcal / g, 1g carbohydrate: 2.5 kcal / g).

Table 2. Amino acid composition in natural feed and the body of sea horse juveniles *H. barbouri* (%) body protein

Amino Acid	Feed treatments					Sea horse juveniles
	A	B	C	D	E	
Essential A. Acid						
<i>Histidine</i>	0.14	0.13	0.12	0.11	0.11	0.12
<i>Arginine</i>	0.09	0.09	0.08	0.07	0.06	0.28
<i>Threonin</i>	0.24	0.20	0.16	0.13	0.09	0.19
<i>Valine</i>	0.32	0.29	0.27	0.24	0.21	0.27
<i>I-leucine</i>	0.28	0.25	0.22	0.20	0.17	0.17
<i>Leucine</i>	0.40	0.36	0.33	0.30	0.27	0.30
<i>Lysine</i>	0.24	0.22	0.21	0.19	0.18	0.20
<i>Phenylalanine</i>	0.23	0.21	0.20	0.18	0.17	0.27
<i>Methionine</i>	0.47	0.39	0.30	0.22	0.13	0.18
<i>Tryptophan</i>	0.05	0.04	0.03	0.02	0.01	0.03
Non-Essential A. Acid						
<i>Aspartic acid</i>	0.23	0.21	0.19	0.17	0.15	0.29
<i>Glutamic acid</i>	0.75	0.68	0.62	0.56	0.50	0.48
<i>Serine</i>	0.21	0.19	0.16	0.14	0.11	0.18
<i>Glycine</i>	0.33	0.34	0.35	0.37	0.38	0.58
<i>Alanin</i>	0.38	0.38	0.38	0.38	0.38	0.49
<i>Proline</i>	0.28	0.22	0.17	0.12	0.07	0.32
<i>Tyrosine</i>	0.08	0.07	0.06	0.05	0.05	0.12
<i>Sistine</i>	<0.004	<0.004	<0.004	<0.004	<0.004	<0.004

Notes :

1. A(100% naupli *Artemia*), B(75% naupli *Artemia*+25% *Phronima*), C(50% naupli

- Artemia*+50% *Phronima*), D(25% nauplii *Artemia*+75% *Phronima*) dan E(100% *Phronima*);
- The value of amino acid in the juvenile body is in A and E treatments were obtained from the result of analysis in Laboratorium;
 - The values of amino acid in B, C, and D treatments were obtained from calculation.

Amino acid analysis produced, ten essential amino acid content found from the body of juvenile seahorse i.e., arginine, lysine, leucine, isoleucine, phenylalanine, valine, methionine, histidine, threonine, and tryptophan (Table 2). The results of the amino acid analysis showed that several types of essential amino acids were deficient in the seahorse juvenile requirements. The number of essential amino acids arginine and phenylalanine in all treatments A, B, C, D, and E were known deficient in juvenile requirements. C treatment deficient in threonine. D treatment was deficient in histidine, threonine, valine, lysine, and tryptophan

In treatment E, the total overall essential amino acid contained deficient in juvenile needs.

Survival rate

Table 3. The mean of survival rate (%) of sea horses juveniles *Hippocampus barbouri*

Treatments	Survival rate (%)
A. 100% <i>Artemia</i> naupli	80.00 ± 10.00 ^{ab}
B. 75% <i>Artemia</i> naupli + 25% <i>Phronima</i> sp.	90.00 ± 17.32 ^{ab}
C. 50% <i>Artemia</i> naupli + 50% <i>Phronima</i> sp.	96.67 ± 5.77 ^b
D. 25% <i>Artemia</i> naupli + 75% <i>Phronima</i> sp.	96.67 ± 5.77 ^b
E. 100% <i>Phronima</i> sp.	63.33 ± 15.27 ^a

Notes: Different letters indicated a significant difference (P<0.05) of survival rate of juveniles

Substitution of *Artemia* naupli with *Phronima* sp. up to 75% in treatment D showed a survival rate that was not different from giving 100% *Artemia* naupli in treatment A (Table 3). The resulting synthesis in treatment D is 96.67 ± 5.77% while in treatment A is 80.00 ± 10.00 %. However, the substitution of naupli *Artemia* with *Phronima* up to 100% in treatment E has not been given to the maintenance of seaweed juveniles *H. barbouri*, which is characterized by the low survival value produced.

The survival of juveniles is influenced by several important factors, one of which is the adequacy of feed nutrients provided during maintenance. Adequacy of juvenile nutrition was analyzed based on proximate analysis and amino acid. The nutritional content of the proximate analysis of the substitution of *Artemia* and *Phronima* nauplii in treatment B, C, and D is sufficient for the number of juvenile needs, especially in the protein and fat

content. The protein and fat content in treatment B was 55.38% and 17.12%; treatment C 49.30% and 12.69% and treatment D 43.21% and 8.25%. Juvenile protein and fat requirements are known to range from 40-60% and 10-20% respectively (Morais *et al.*, 2001; Watanabe *et al.*, 2001; Skalli *et al.*, 2004 In Woods, 2007). The initial stage of juvenile growth is known to require high protein content (Nenciu *et al.*, 2015; Novelli *et al.*, 2016). The availability of protein and fat in the feed must be at the optimum level because it will affect the work system of the body's metabolism of larvae or juveniles. The protein and fat content is the primary fuel source that determines the quality of the juvenile's ability to survive (Blanco *et al.*, 2011).

The results of the amino acid substitution analysis of *Artemia* naupli with *Phronima* in treatments A, B, and C was sufficient from the average number

of essential amino acids needed for the survival of juveniles. The quality of the protein content of a feed can be seen from the adequacy of essential amino acids in the body's juvenile needs. The existence of amino acids provides different functions for the growth and development of an organism. Amino acids are important basic ingredients during the process of larval growth (Ronnestad *et al.*, 2003). Essential amino acids that are known to play an important role in increasing larval growth and development, namely in histidine, valine, and lysine (Buwono, 2000; Li *et al.*, 2008).

Winarno (2008) explains that amino acids needed by fish must be fulfilled from the feed, especially for amino acids that cannot be synthesized (essential) by the body. Amino acids are the main constituent components of proteins and are divided into two groups, namely essential and non-essential amino acids. Essential amino acids cannot be produced in the body so they often have to be added in the form of food, while non-essential amino acids can be produced in the body (Sitompul, 2004). Essential amino acids are an important source of energy sources to ensure adequate protein content of feed during the growth period (Ronnestad *et al.*, 2003). According to Wijaya (2003), amino acids are needed by the body as an energy source and can function as a source of material for protein synthesis which is very much needed by larvae or juveniles during the organ formation phase.

The low survival rate of substitution of *Artemia* naupli with *Phronima* up to 100% treatment E is thought to be due to the low protein and fat content of the number of juvenile requirements, as well as the amino acid content. Also, the total energy

produced by treatment E is low from other treatments (A, B, C, and D). The total energy in treatment A 579.27 kcal/g; treatment B 497.32%; treatment C 415.36% while treatment E 251.45%. Foods with low energy content can cause fish to use some protein as an energy source for metabolism so that the protein portion for growth is reduced (Pramono *et al.*, 2007). Besides that, the high crude fiber content in treatment E also causes low growth and survival in the maintenance of seahorse juveniles belonging to carnivorous species. According to Afrianto & Liviawati (2005), the ability to digest carnivorous fish against crude fiber is relatively low compared to fat and protein. The higher the crude fiber, the higher the feed is difficult to digest (Satyani, 2003) so that the growth and survival of the juvenile become low.

During maintenance of juveniles, the environmental conditions of maintenance media are at the optimum tolerance and growth limits for seaweed *H. barbouri* juveniles. Temperature 28-30 (Lin *et al.*, 2009; Abidin *et al.*, 2008), pH 6-7 (Abidin *et al.*, 2008; Syafiuddin, 2008), salinity 30-31 ppm (Abidin *et al.*, 2008), DO 4, 2-6 mg/L and ammonia <0.01 mg/L (Abidin *et al.*, 2008).

CONCLUSION

Based on the analysis of the quality of seaweed juwana *H. barbouri* substitution of *Artemia* naupli with *Phronima* sp. up to 75% in treatment D gives a survival value that is not different from giving 100% *Artemia* naupli in treatment A. Therefore, the use of *Phronima* sp. as an alternative feed for the maintenance of seahorse juveniles can be given at a substitution rate of up to 75%.

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